

CENTER FOR RECONSTRUCTIVE URETHRAL SURGERY



GUIDO BARBAGLI, M.D.
Arezzo - Italy

e-mail: info@urethralcenter.it

Websites: www.uretra.it
www.urethralcenter.it



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CENTER FOR GENITO-URETHRAL
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BELGRADE, FEBRUARY 23-26, 2015

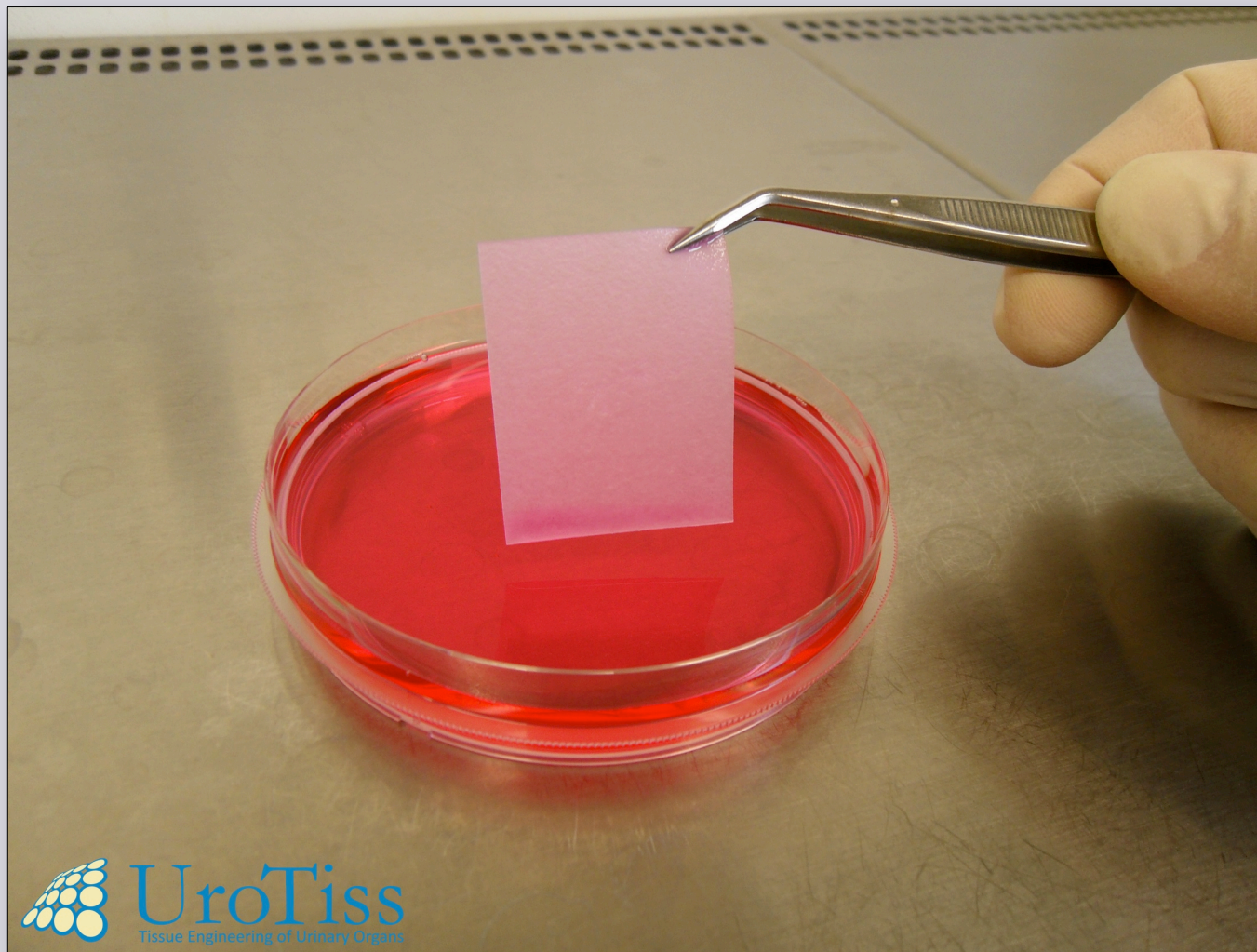
1ST INTERNATIONAL COURSE ON URO-GENITAL RECONSTRUCTIVE SURGERY

— LIVE SURGERY AND LECTURES —

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Tissue engineering urethroplasty



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The Lancet

The end of the beginning for tissue engineering



For three decades we have heard about the hope of tissue engineering. Hyperbole has become routine, but amidst unreasonable expectations are serious scientists, like Paulo Macchiarini, who believe that by combining cells and degradable materials *ex vivo* they can develop organs to replace or repair diseased tissues. After many years of trying to build engineered tissues on a backbone of synthetic degradable polymers,¹ a growing body of evidence suggests that decellularised whole organs and tissues are clinically effective degradable scaffolds.² Until recently, decellularised tissues were used clinically without the addition of cells, and in many cases—eg, the decellularised porcine small intestine submucosa family of surgical implants—this approach was sufficient to generate a healing response. The idea of whole organ engineering—whereby organs are decellularised and then repopulated with desired mixtures of cells—seems to be a realistic path towards complex three-dimensional tissue engineering.

In 2008, Macchiarini's team announced that they had successfully grown a neo-trachea from a decellularised

evidence that the tracheal graft is now naturalised. More importantly, given the data for extracellular-matrix-derived restorative degradable materials and their use in airway and bladder neo-organ development^{4,5} we can celebrate the end of the beginning for tissue engineering; the groundwork has been laid for clinical implementation in other specialties.

Excitement about tracheal regenerative therapy might be muted by realisation that the patient in this study was not restored to full health. Although heroic in complying with the needs of a research study, the patient is suffering from ongoing complications from scarring at the proximal anastomotic site. There is nothing unusual about a tracheal stricture forming at a surgical site and, in fact, this patient had already had such a post-surgical stricture. Rather, the formation of a stricture shows that the remaining challenges for tissue engineering of thin hollow organs such as trachea, oesophagus, intestine, blood vessels, and bladder relate to how neo-tissues are incorporated into existing structures. Research is needed to understand how to



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[http://dx.doi.org/10.1016/S0140-6736\(13\)62033-4](http://dx.doi.org/10.1016/S0140-6736(13)62033-4)

Tissue engineering's green shoots of disruptive innovation



The ready availability of tissues or organs to replace or repair those diseased or damaged is a ubiquitous clinical need, and the rapidly developing field of tissue engineering might offer innovative solutions. Two Articles^{1,2} in *The Lancet* show the incremental expansion of the applications of tissue-engineering technology to reconstructive surgery.

Application of cells to a scaffold, with or without added chemical or mechanical stimuli, followed by their use for repairing congenital or acquired defects was well established in the 1990s³ and has led notably to the treatment for congenital bladder defects clinically available in the USA. Following on well documented successes in the clinical application of tissue-engineered skin,⁴ blood vessels,⁵ urethra,⁶ and bladder wall,⁷ investigators used similar principles of seeding autologous differentiated cells onto collagen scaffolds to treat patients with vaginal aplasia.¹

In four girls aged 13–18 years, with a rare form of vaginal aplasia, Atlántida Raya-Rivera's team

restoration of contour and nasal airflow to the noses of two women and three men, aged 76–88 years, undergoing substantial resections of external nasal tissue as treatment for skin cancer. After flap refinement at 6 months, Fulco and colleagues² took biopsy samples of repair tissues and histologically analysed them. Safety and feasibility of the procedure 12 months after reconstruction were the primary outcomes. Importantly, the staged reconstruction in the patients permitted histological assessment of the implanted tissue and confirmed sustained restoration of all three layers of the nose that had been reconstructed. At 12 months, no adverse events had been recorded and patients were satisfied with the aesthetic and functional outcomes.

Together, these two studies^{1,2} begin to answer three of the key scientific questions posed to, and by, the translational tissue-engineering community.⁸ First, are biological scaffolds, with or without cells, replaced by scar tissue or native quality tissue over time? In both studies, findings show excellent evidence of multilayer remodelling in a manner consistent with the normal tissues restored. Second, can tissue-engineered



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[http://dx.doi.org/10.1016/S0140-6736\(14\)60544-4](http://dx.doi.org/10.1016/S0140-6736(14)60544-4) and
[http://dx.doi.org/10.1016/S0140-6736\(14\)60542-0](http://dx.doi.org/10.1016/S0140-6736(14)60542-0)

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The Lancet, April 2014

e-mail: info@urethralcenter.it

Websites: www.uretra.it
www.urethralcenter.it

Repair of the anterior urethra is one of the most demanding surgical problems in urology. In recent years, regenerative medicine and tissue engineering studies have led to the development of novel biomaterials for urethral repair.

Here we describe the first clinical report of a series of patients who underwent tissue-engineered oral mucosal graft urethroplasty for anterior urethral stricture.

Dr. Gouya Ram-Liebig and Dr. Soeren Liebig
UroTiss GmbH – Dresden - Germany



Prof. G. Barbagli - Dr. G. Romano – Dr. M. Lazzeri
Center for Reconstructive Urethral Surgery – Arezzo - Italy

e-mail: info@urethralcenter.it

Websites: www.uretra.it
www.urethralcenter.it



Dresden - Germany

UroTiss GmbH is a pharmaceutical company, founded in Germany in 2005 by Dr. Gouya Ram-Liebig and Dr. Soeren Liebig. UroTiss provides products with highest safety and quality, in accordance to current Good Manufacturing Practices (GMP).

www.urotiss.com

Email: g.ram-liebig@urotiss.com

e-mail: info@urethralcenter.it

**Websites: www.uretra.it
www.urethralcenter.it**

Preclinical and clinical examination of tissue-engineered graft for urethral reconstruction (MukoCell®) with regard to its safety

Massimo Lazzeri¹, Guido Barbagli¹, Dirk Fahlenkamp², Giuseppe Romano³, Ulf Balsmeyer², Helmut Knispel⁴, Maria-Elsa Spiegel⁴, Burkard Stuerzebecher⁴, and Gouya Ram-Liebig⁵

(1) 1 Centre for Reconstructive Urethral Surgery, Arezzo, Italy (2) Zeisigwald Clinics Bethanien, Department of Urology, Chemnitz, Germany (3) San Donato Hospital, Department of Urology, Arezzo, Italy (4) St. Hedwig Krankenhaus, Department of Urology, Berlin, Germany (5) UroTiss GmbH, Dresden, Germany

I. Introduction

MukoCell® is a national authorized, autologous tissue-engineered oral mucosa graft. The present report sums up some of MukoCell®'s preclinical safety data. Additional reported data of 70 patients, treated with MukoCell®, are also considered with regards to safety analysis.

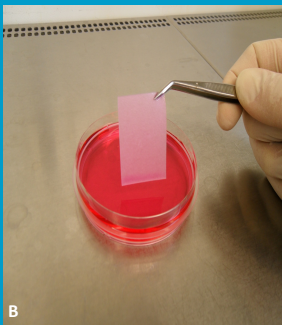
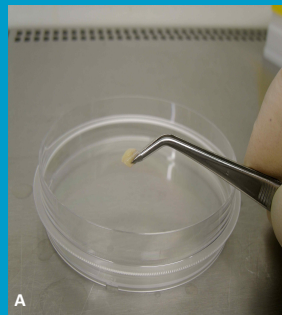


Fig. 1: Patient's oral mucosa cells are generated from a small oral mucosa biopsy (A) and cultured on the surface of a biocompatible scaffold (B).

II. Methods

For MukoCell® production, patient's oral mucosa cells were generated from a small oral mucosa biopsy and cultured on the surface of a biocompatible scaffold (Fig. 1).

The tumorigenic potential of MukoCell® was examined in vivo. For this purpose, human cultured cells of 4 different runs were injected by intraperitoneal and subcutaneous route into each of ten immunodeficient athymic nude mice. 4×10^7 cells $\pm 2 \times 10^6$ cells were injected into each animal on Days 1, 18, 25 and 46 of the study. An additional group consisting of ten animals each received cell culture medium as vehicle control (Table 1).

To examine the potential migration of cells into distant organs, murine MukoCell® constructs from eGFP-transgenic mice were implanted into peritoneal cavity of histocompatible non-transgenic mice and vice versa. The 24 test animals were sacrificed either at weeks 1, 2, 4 or 12 for histological analysis (Table 2).

To investigate the degradation of implanted MukoCell® with time, scaffolds with the size of 0.5 x 1.5 cm were implanted into the peritoneal cavity of 20 BALBc/C57BL6J mice.

Additionally, reported clinical safety data from 70 MukoCell®-treated patients with urethral stricture, which have been recruited in an ongoing observational study with up to 2 year follow-up period, were evaluated on the basis of a pharmacovigilance system. Ethical committee votum was available for the study.

III. Tables

Table 1. Experimental group allocated during the tumorigenicity study

Group	No. of animals	Item	Injection on days ¹⁾	Injection volume (i.p. + cells at each day s.c.) [μ L]	Total no. of cells
1a	5	Test items (n=4)	1, 18, 25, 46	200 + 200	$10^7 \pm 2 \times 10^6$
1b	5	Test items (n=4)	1, 18, 25, 46	200 + 200	$10^7 \pm 2 \times 10^6$
2a	5	Control item	1, 18, 25, 46	200 + 200	-
2b	5	Control item	1, 18, 25, 46	200 + 200	-

Notice²⁾ Cell preparations generated independently from four different runs were used. A separate cell preparation was used on each day.

Table 2. Allocation and treatment of animals in the biodistribution study

Group (Cage)	No. of animals	Donor for engineered tissue graft	Recipient of engineered tissue graft	Sacrifice after implantation week
A-1 (11/5/0)	3	EGFP-tg	nontg	after 1 week
A-2 (11/6/0)	3	EGFP-tg	nontg	after 2 weeks
A-3 (11/7/0)	3	EGFP-tg	nontg	after 4 weeks
A-4 (11/8/0)	3	EGFP-tg	nontg	after 3 months
reserve animal (11/H3/0)		EGFP-tg	nontg	
B-1 (11/1/9)	3	nontg	EGFP-tg	after 1 week
B-2 (11/2/9)	3	nontg	EGFP-tg	after 2 weeks
B-3 (11/3/9)	3	nontg	EGFP-tg	after 4 weeks
B-4 (11/4/9)	3	nontg	EGFP-tg	after 3 months
reserve animal (11/H1/9)		nontg	EGFP-tg	

Notice: Material used for histology after sacrifice: Brain (cerebrum, cerebellum, brain stem, paraventricular parts); heart; kidneys;

large intestine (caecum, colon, rectum); liver; lung; lymph nodes (mesenteric)

intestine; (duodenum, jejunum, ileum) / peyer plaques; spleen; thymus; transplants (including surrounding tissue)

IV. Results

Evaluation of tumorigenicity study in nude mice did not reveal macroscopic and microscopic malignancies attributable to MukoCell® in 60 different examined tissues and organs. Additionally, migration of the transplanted cells into distant organs was excluded at all examined time intervals after implantation of murine homologue of MukoCell®. While the grafts were still present in all 10 animals 9 days after implantation, 6 of 10 grafts were degraded 40 days after implantation in the remaining 10 animals. Clinical data of 70 with MukoCell® treated patients demonstrated no peri- or post-operative adverse events related to MukoCell®.

V. Conclusion

MukoCell® seems to be a safe graft for urethroplasty for patients with urethral stricture. The graft is degrading within a few weeks and hence avoids complication associated with persistent implants.

MukoCell® is an autologous tissue-engineered oral mucosa graft

AUA 2014

MP9 – Abstract ID: 14-578

Tumorigenic study:

- cultured cells of human donors were injected by intraperitoneal and subcutaneous route into each of ten immunodeficient athymic nude mice.
- 4×10^7 cells $\pm 2 \times 10^6$ cells were injected into each animal on Days 1, 18, 25 and 46 of the study.
- An additional group consisting of ten animals each received cell culture medium as vehicle control

Results: No macroscopic and microscopic malignancies attributable to MukoCell® in 60 different examined tissues and organs.

Biodistribution study:

- Murine MukoCell® constructs from eGFP-transgenic mice were implanted into peritoneal cavity of histocompatible non-transgenic mice and vice versa.
- The 24 test animals were sacrificed either at weeks 1, 2, 4 or 12 for histological analysis

Results: No migration of the transplanted cells into distant organs.

Degradatin study:

- 0.5 x 1.5 cm MukoCell® scaffolds were implanted into the peritoneal cavity of 20 female BALBc/C57BL6J mice

Results: 60% of the grafts were degraded 40 days after implantation.

Clinical observational study:

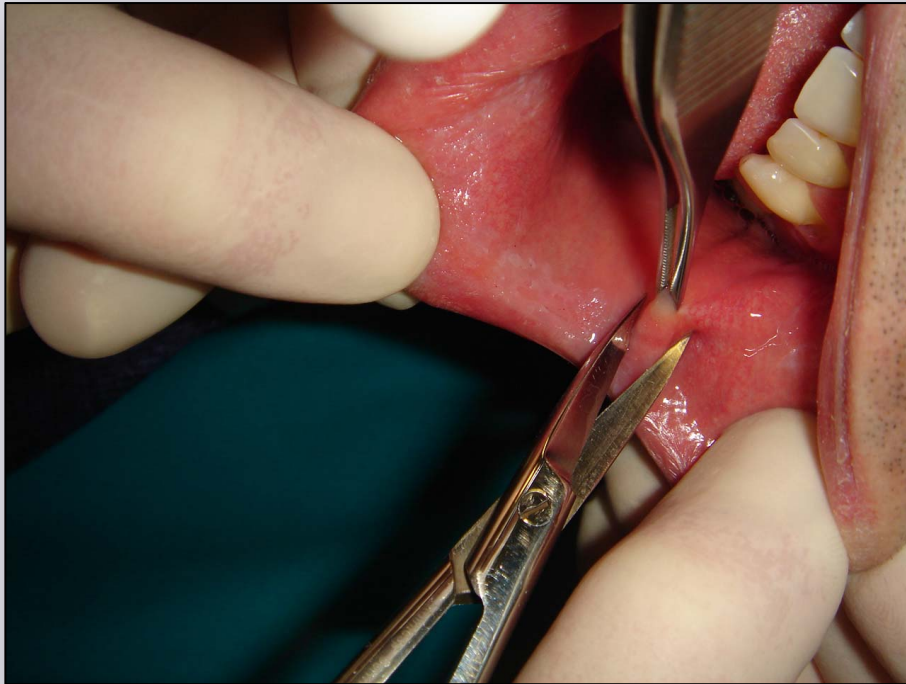
- data from 70 MukoCell®-treated patients with urethral stricture, with up to 2 year follow-up period, were evaluated

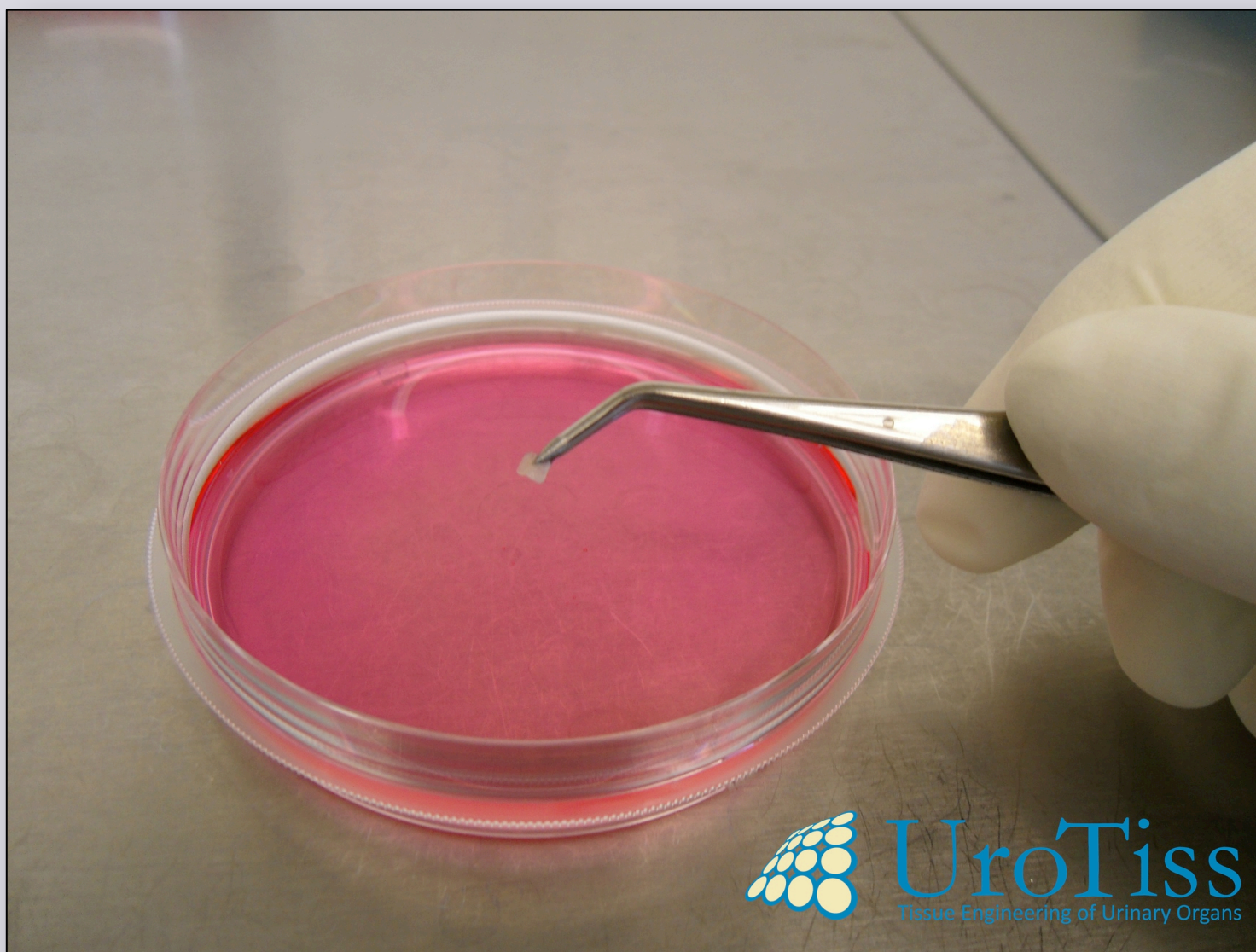
Results: No peri- or post-operative adverse events related to MukoCell®.

Step 1

Harvesting sample from the cheek

Local anaesthesia





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Step 2

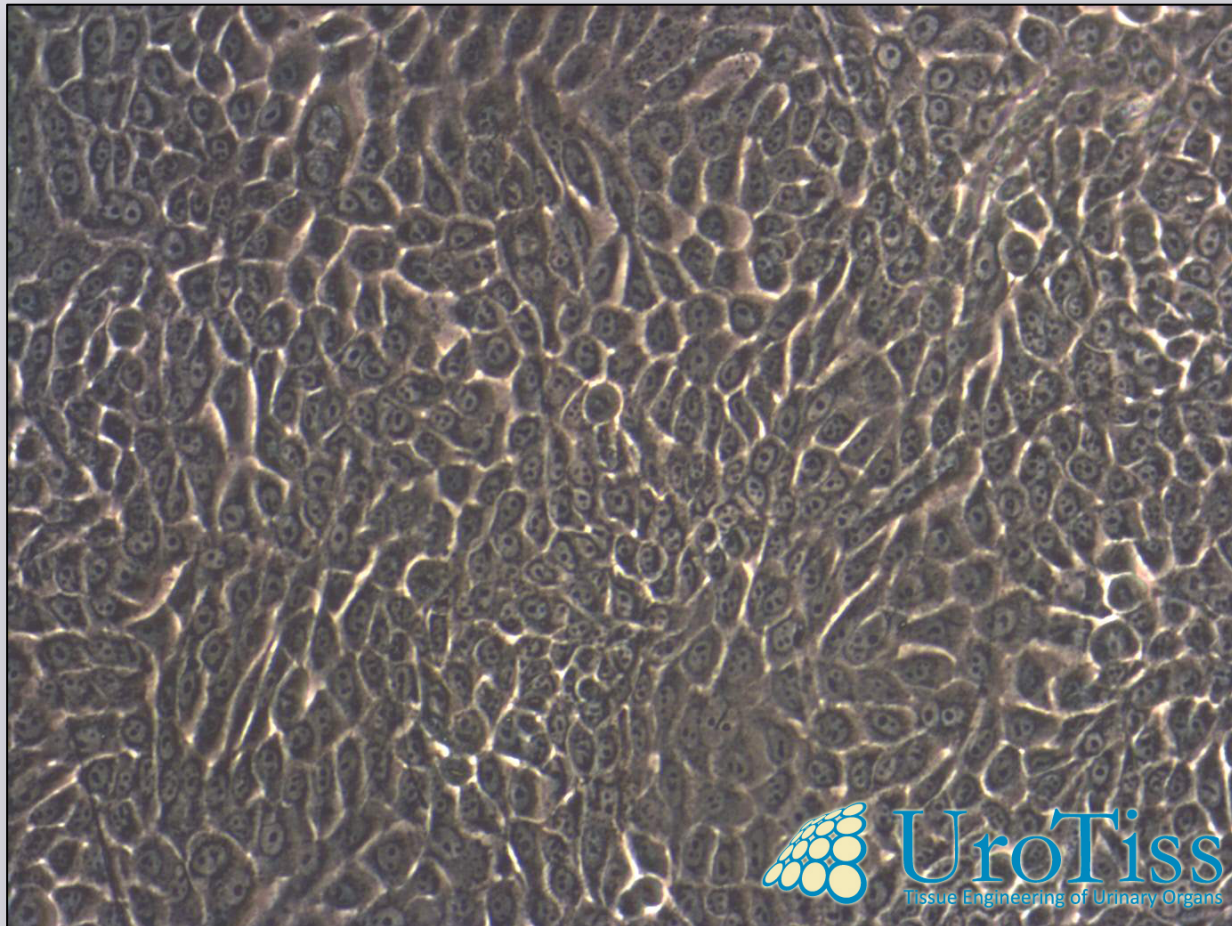
GMP Laboratory



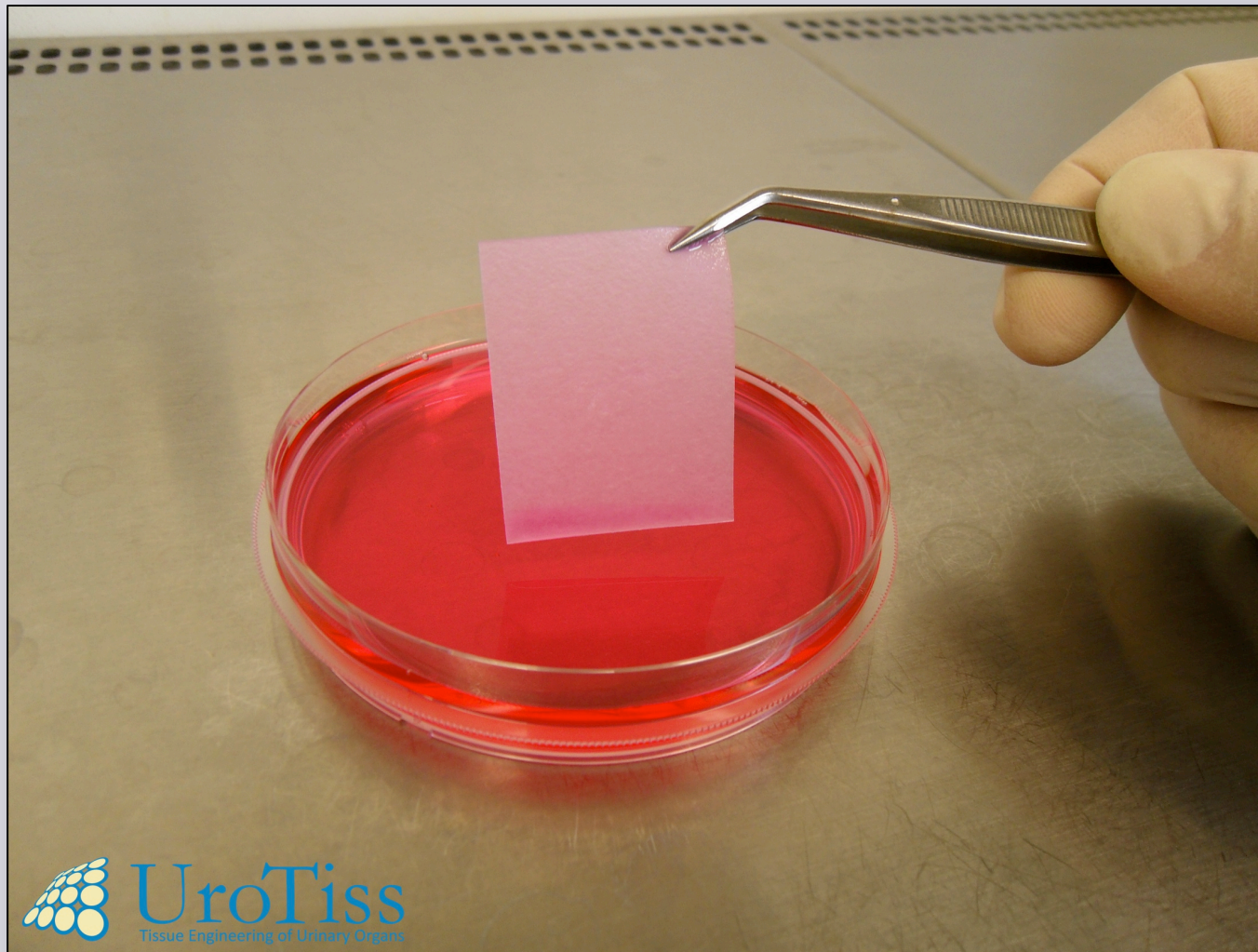
**Clean roon Laboratory in accordance to current
Good Manufacturing Practices (GMP).**

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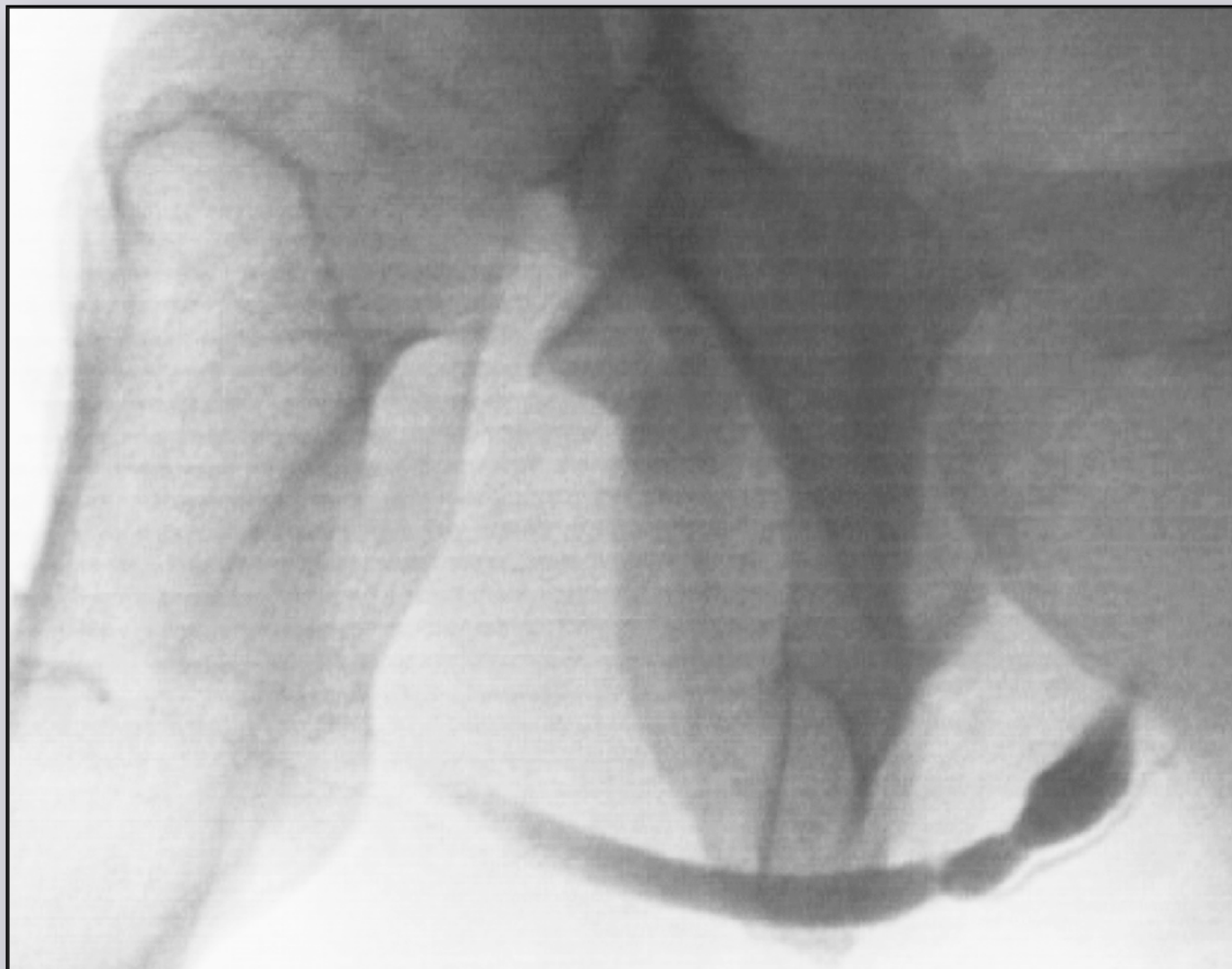


**Cells were expanded
and cultured on the surface of a biocompatible scaffold.**



3 weeks later

Pre-operative retrograde urethrography



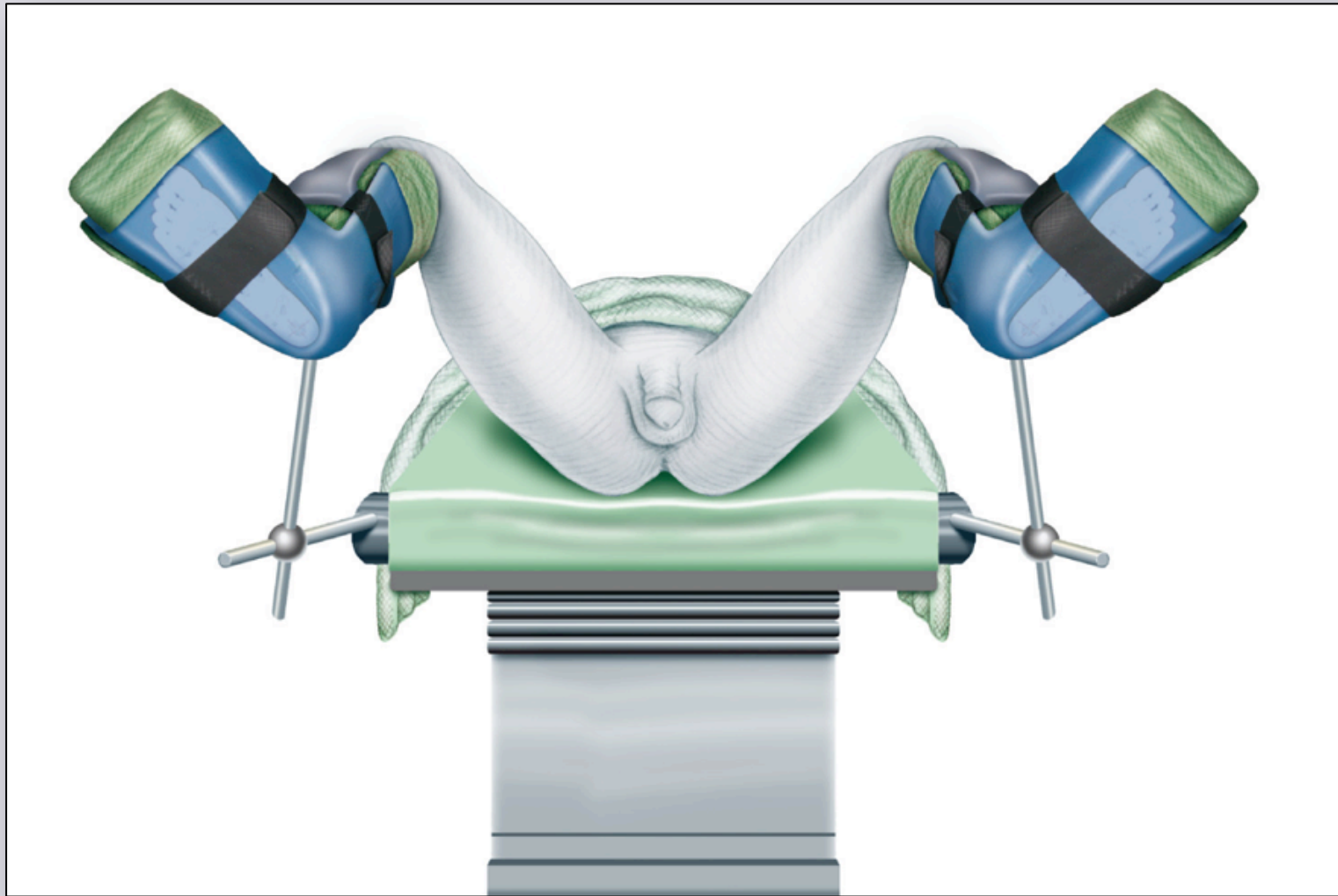
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Step 3

Surgical transplant using dorsal inlay technique



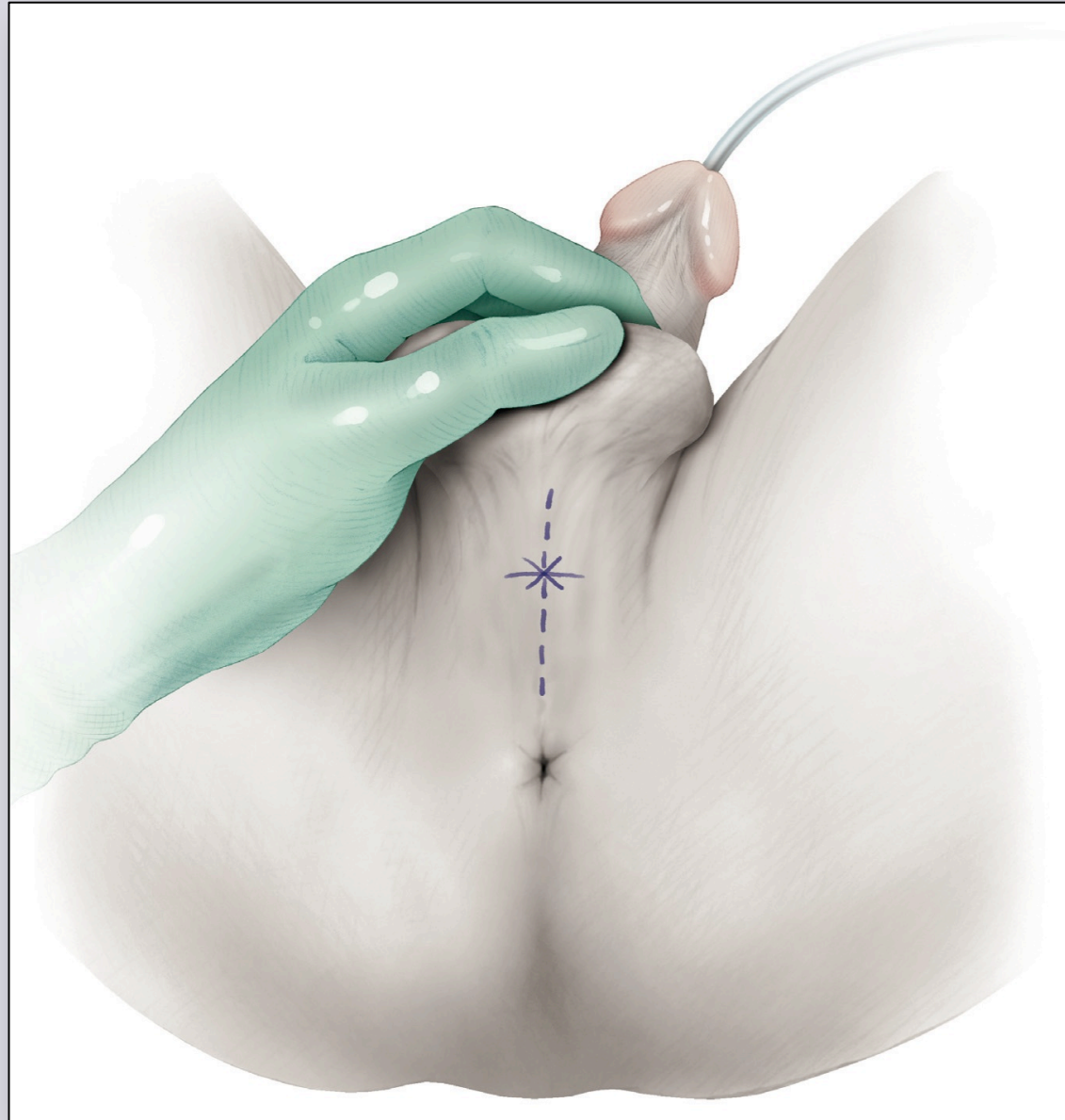
Simple lithotomy position



**Pre-operative
urethroscopy**

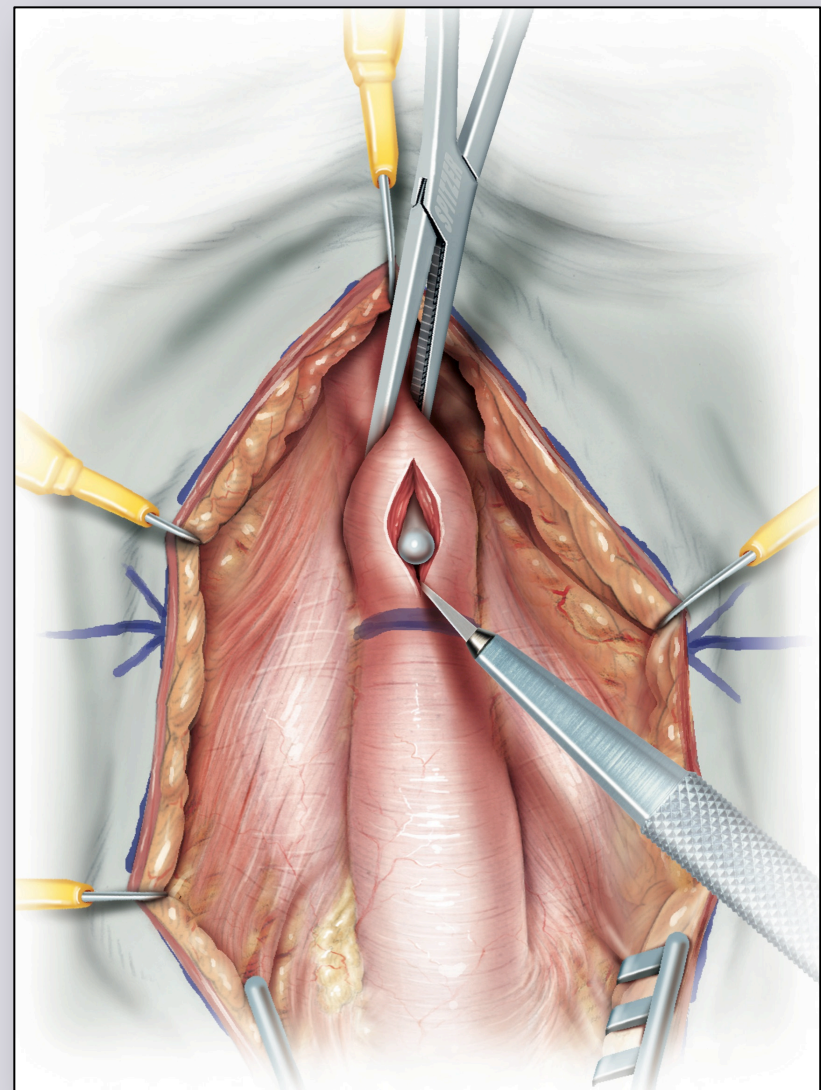
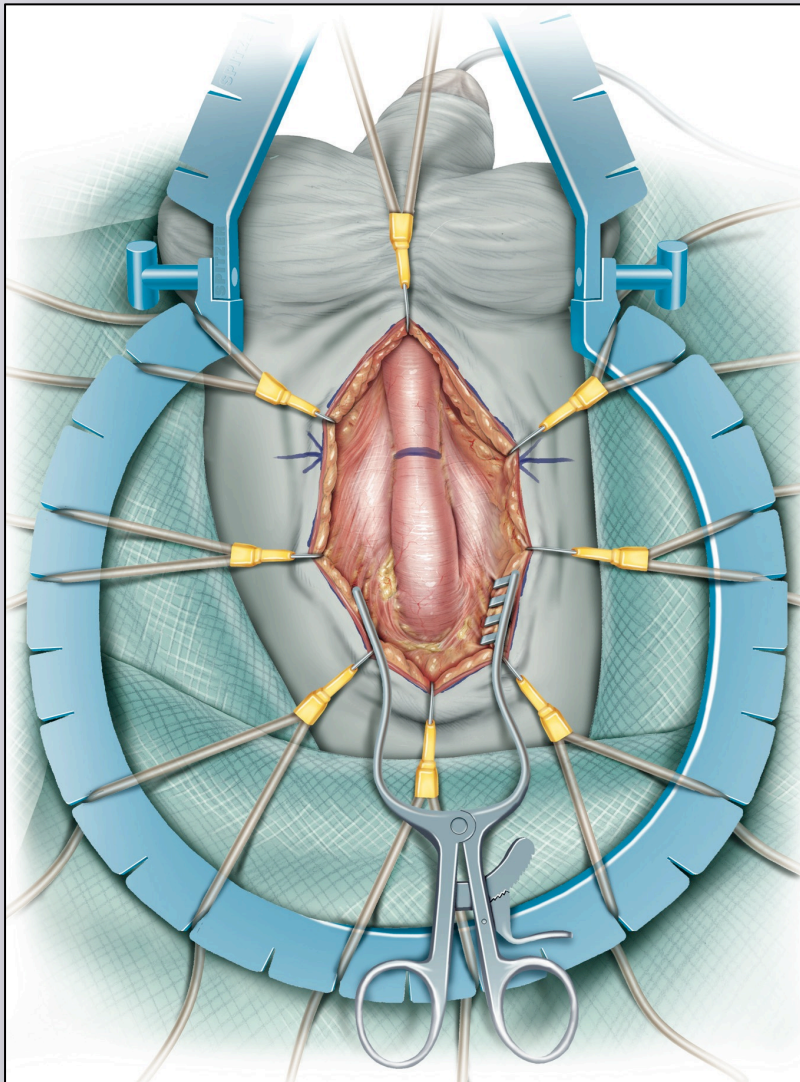


Insert Sensor 3 Fr. guidewire



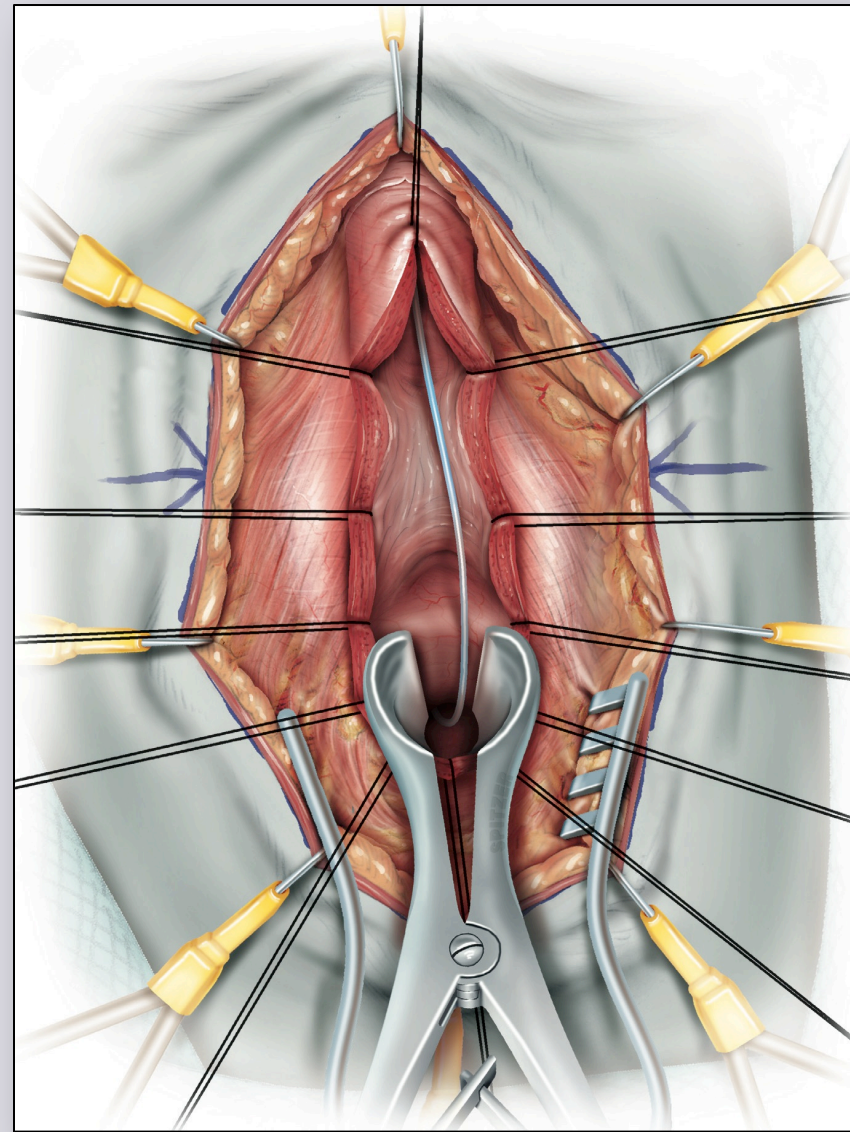
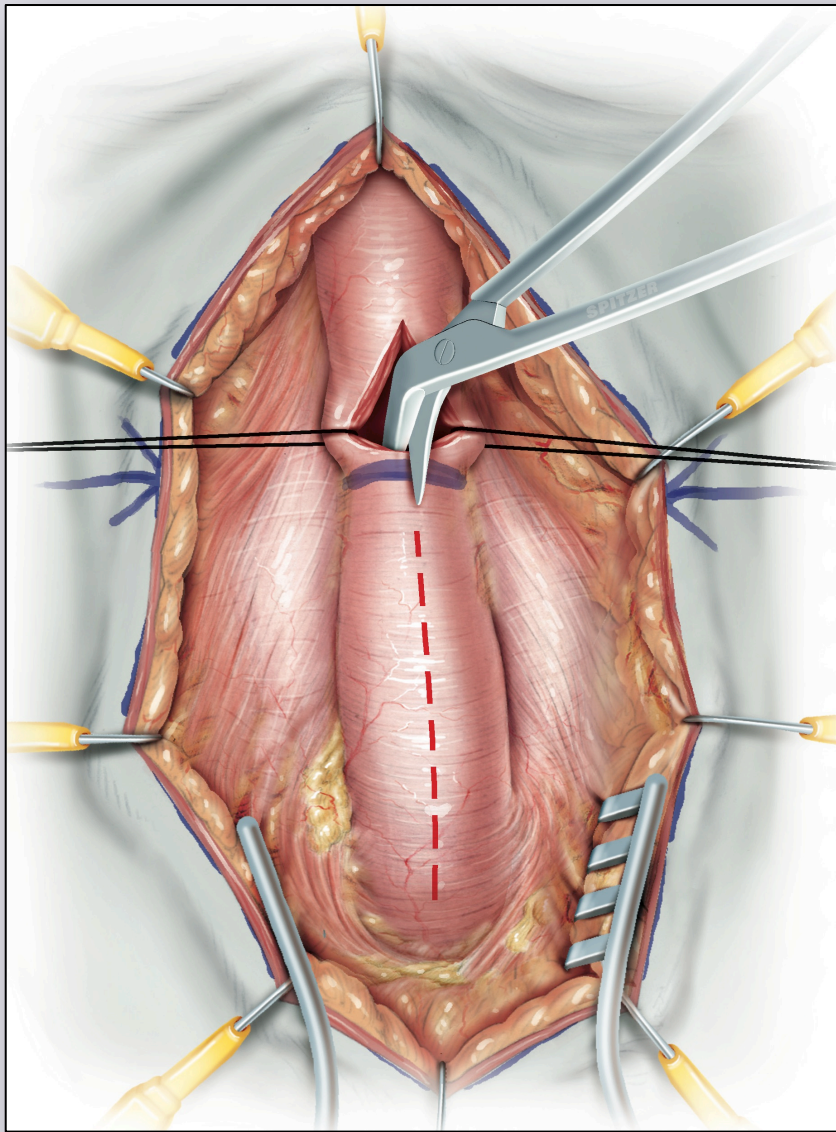
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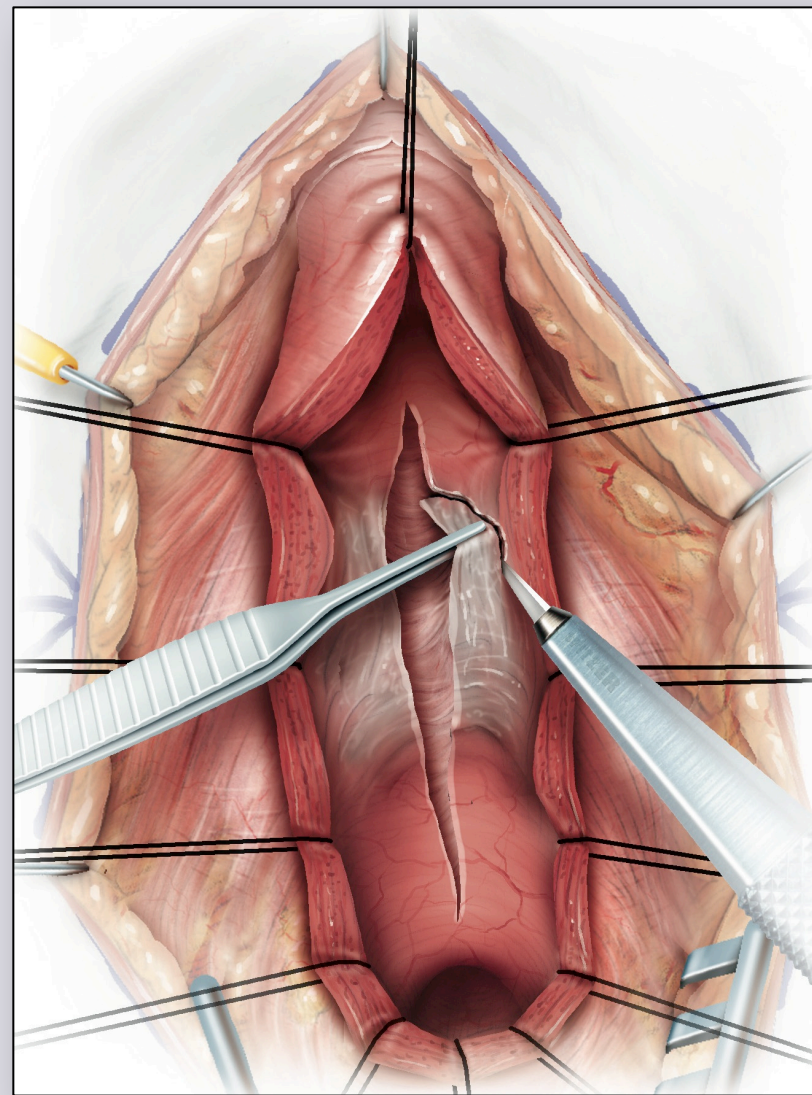
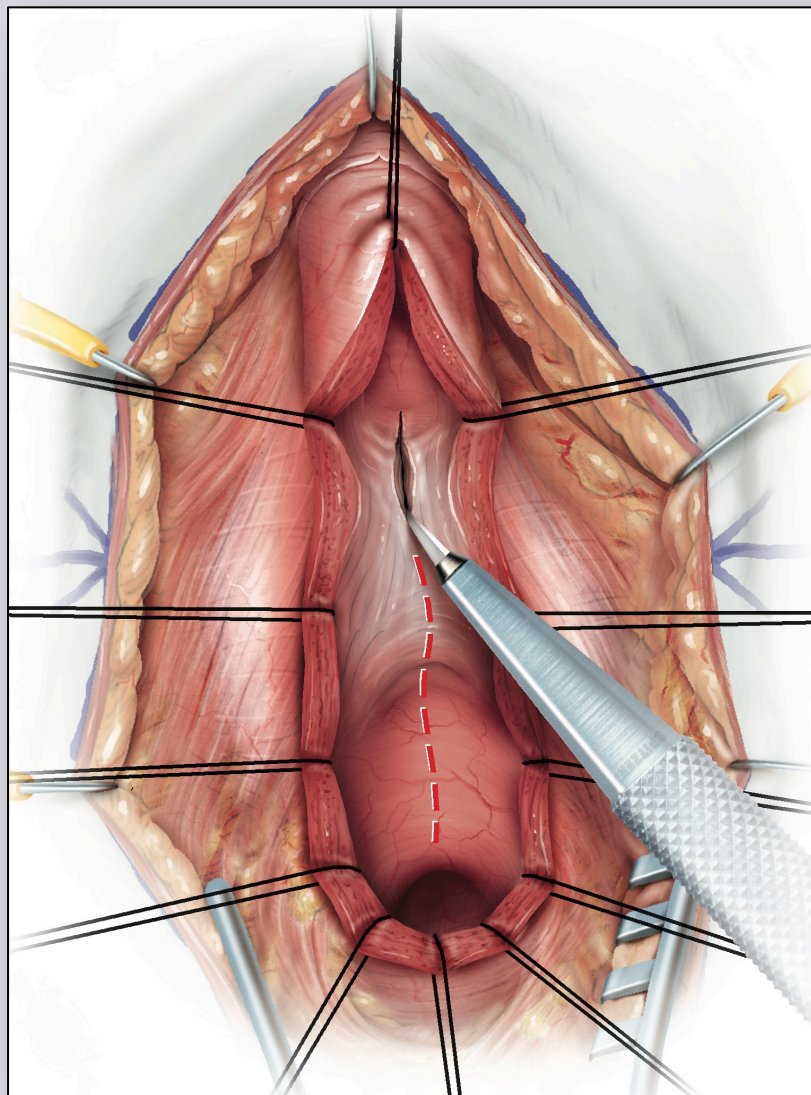
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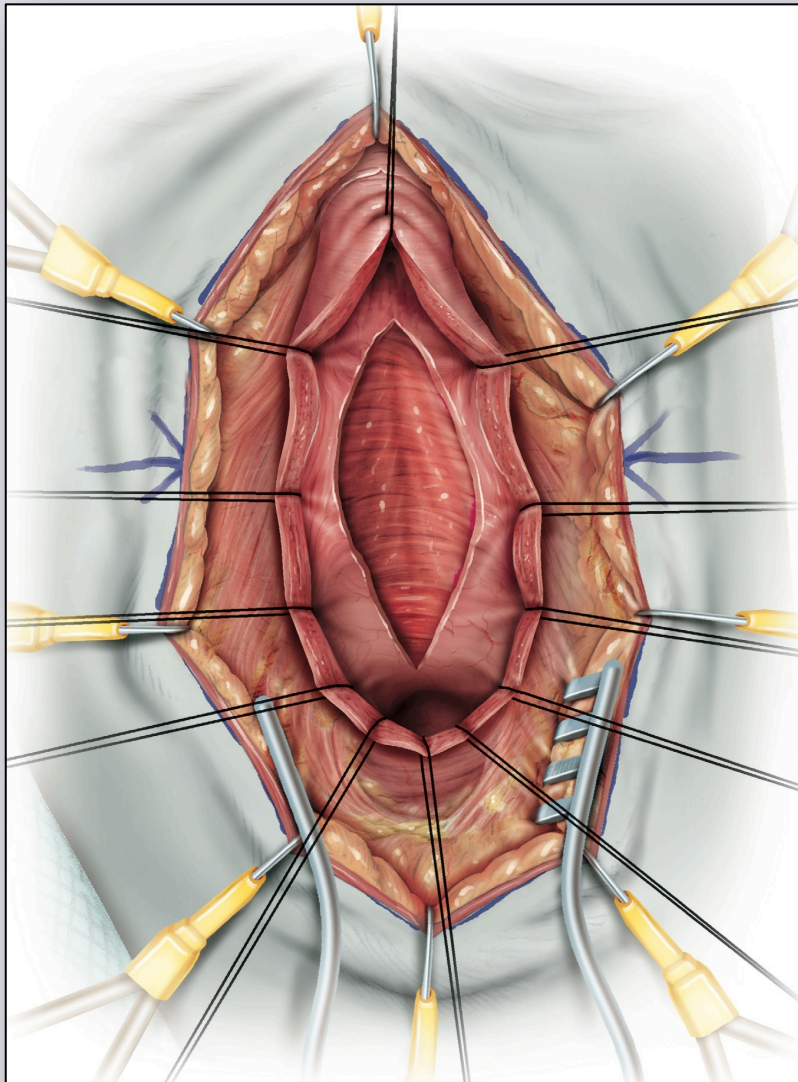
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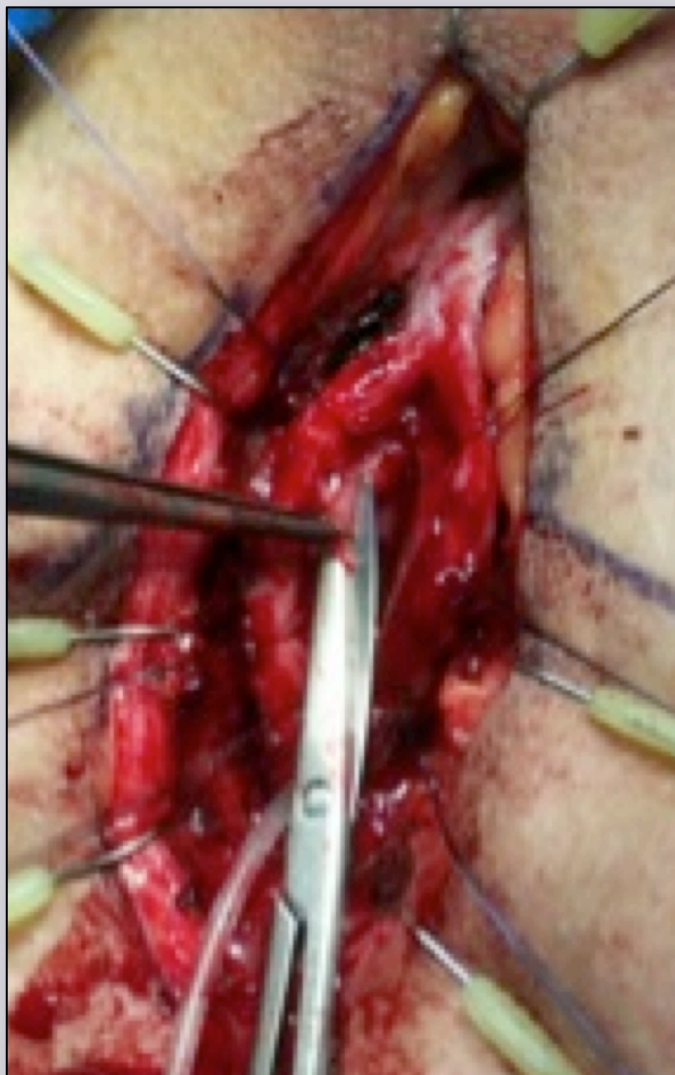
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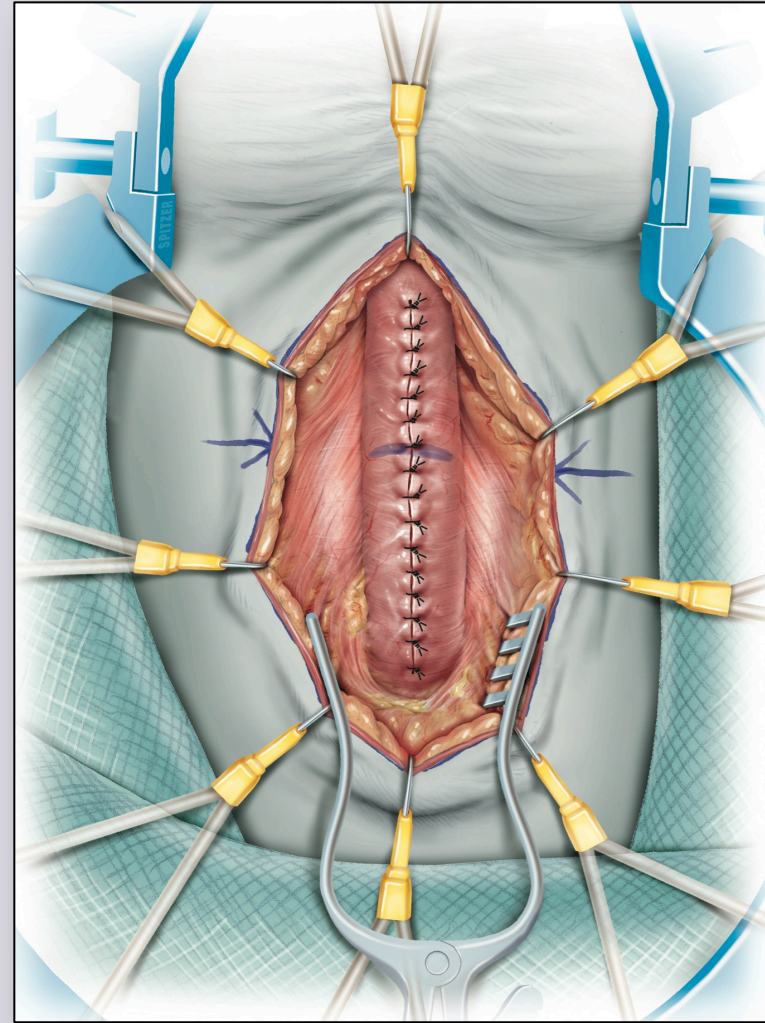
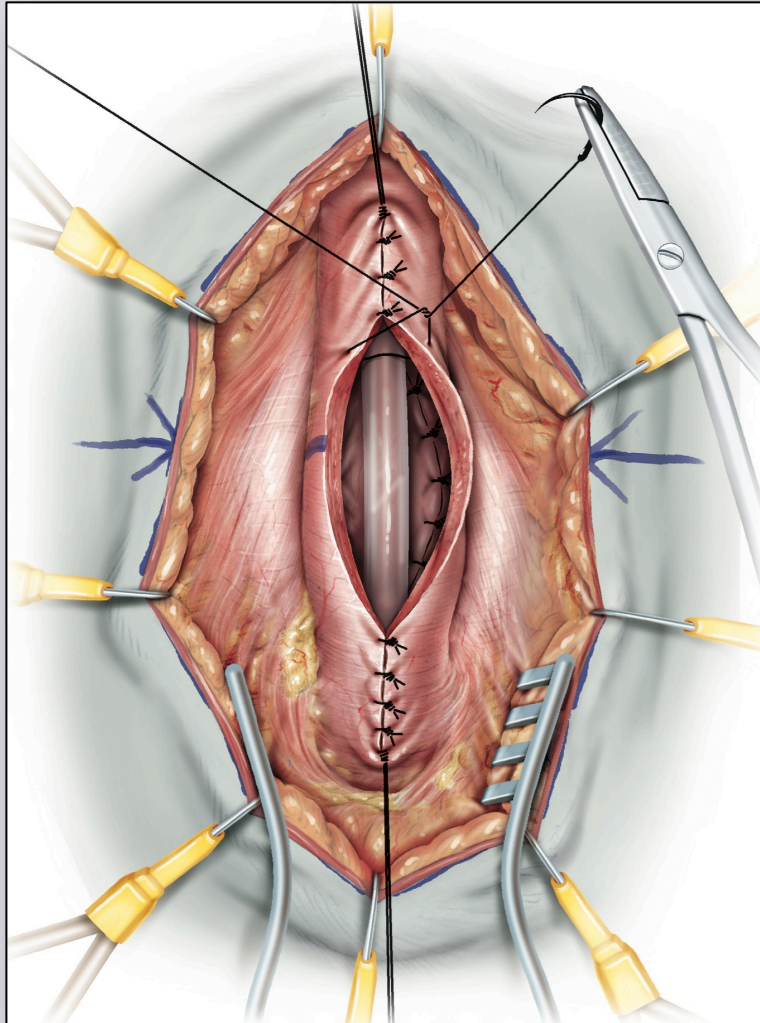
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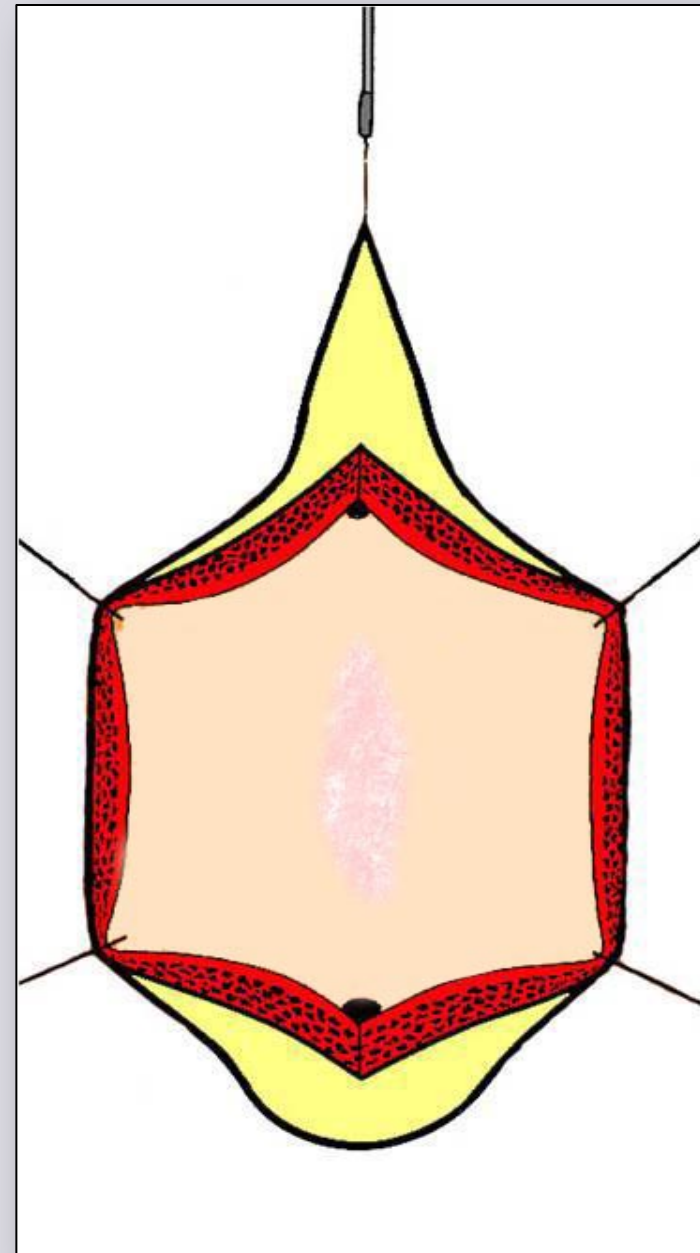
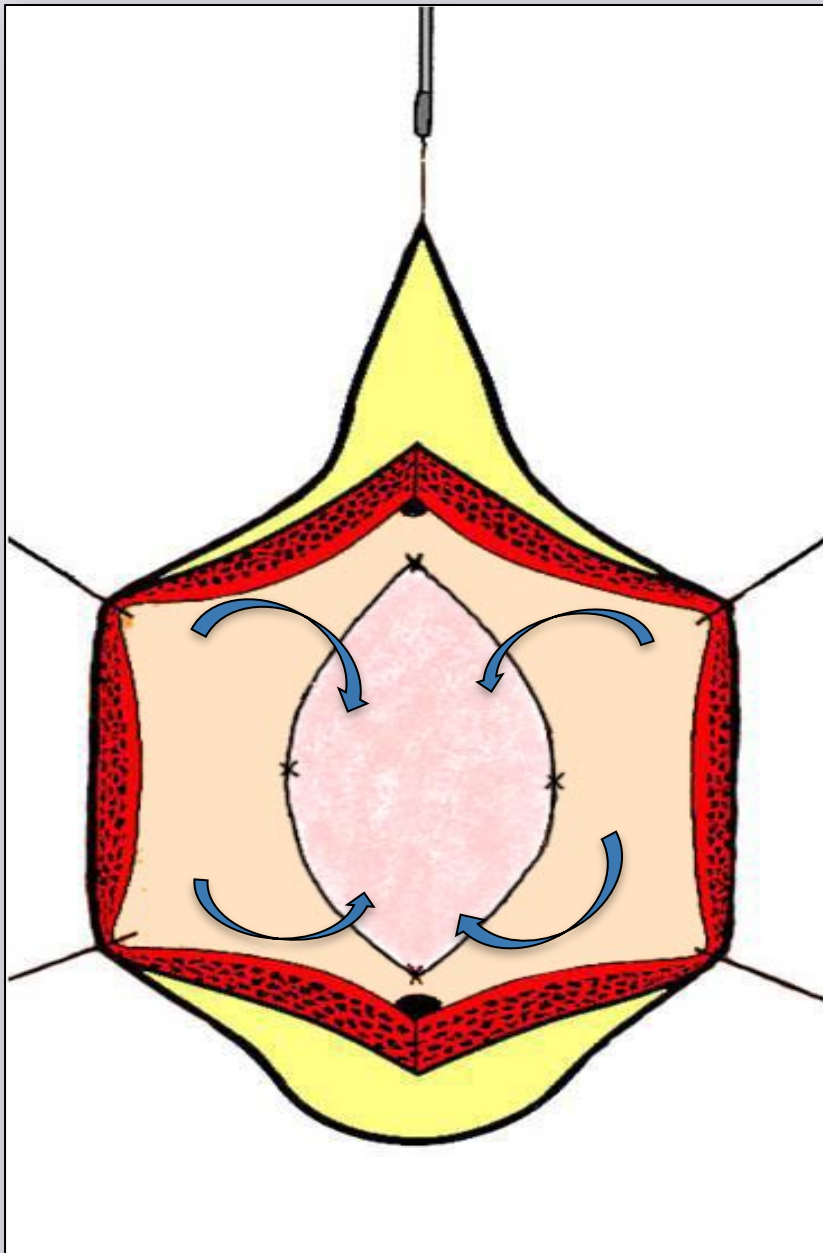
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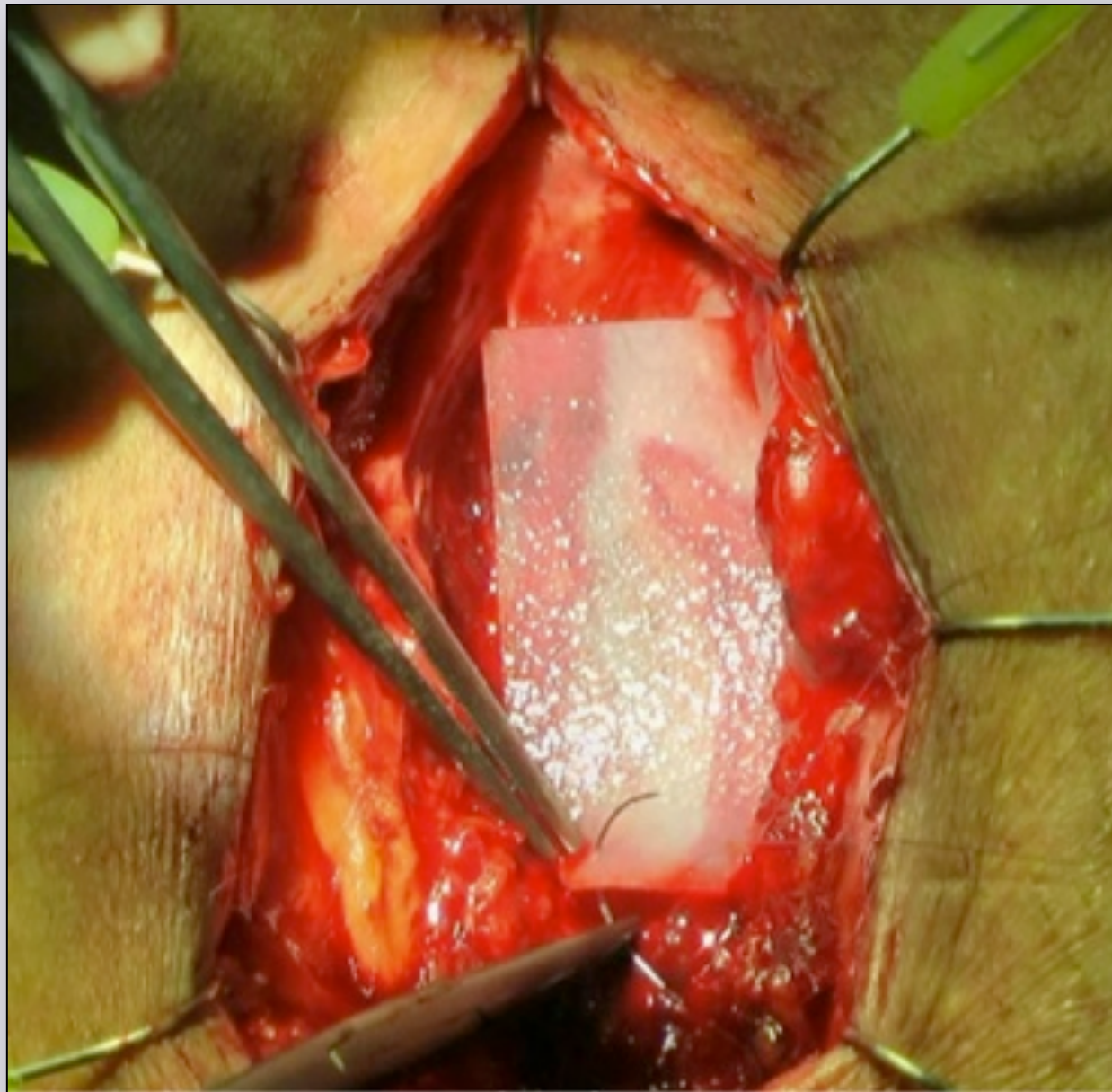
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Surgical transplant using ventral onlay technique



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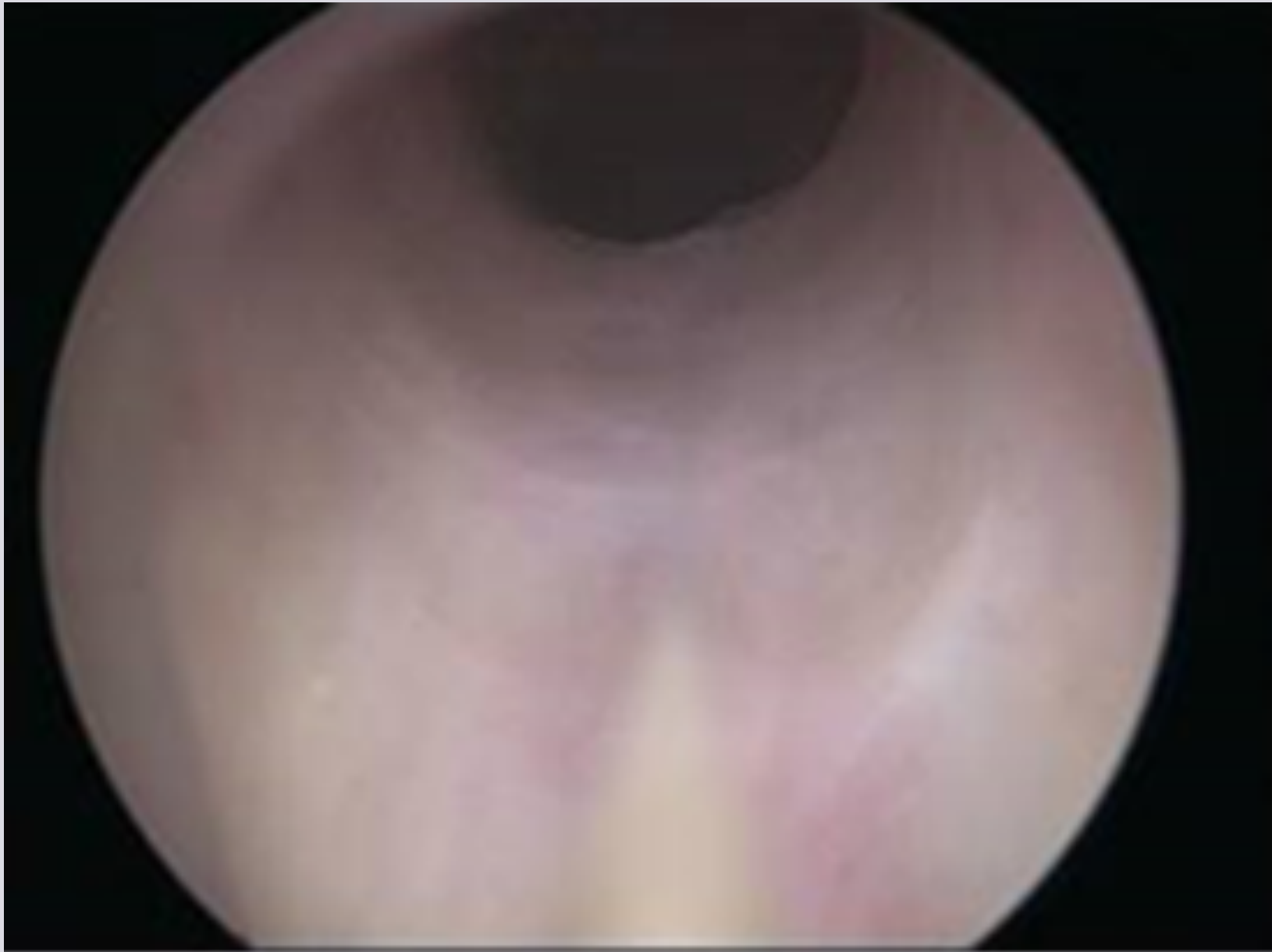
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Post-operative voiding cysto-urethrography



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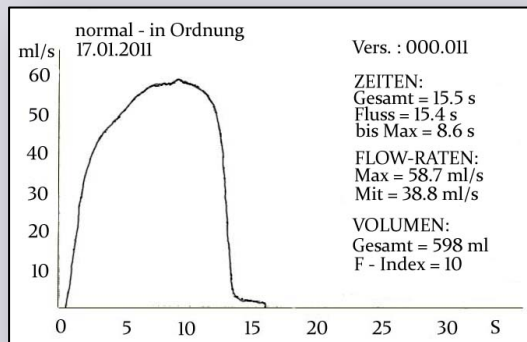
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Clinical experience with MuKocell implant in Germany

Osnabruck
Chemnitz
Berlin
Hamburg
Luneburg
Lipsia

103 patients

Period: from 2010 to 2013

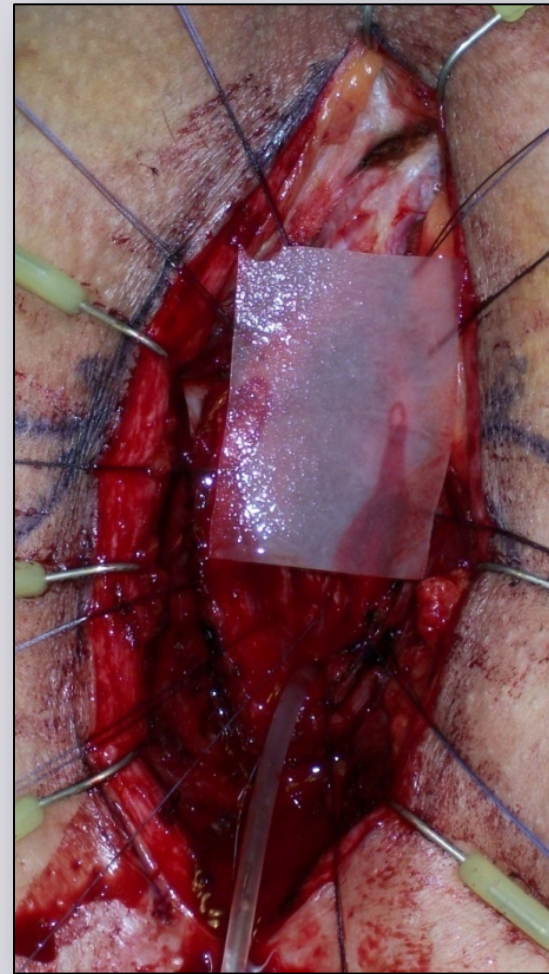


Overall success rate:
from 82% to 85%

Our preliminary experience



Oral mucosa



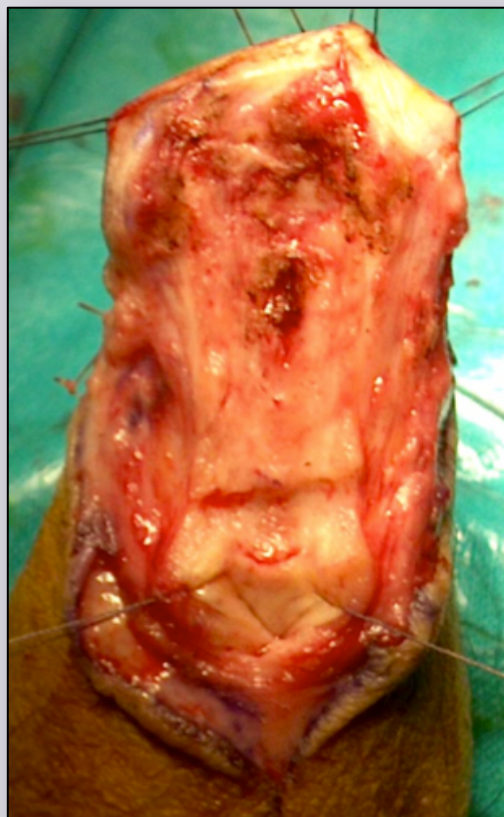
Tissue engineered oral mucosa

Our preliminary experience



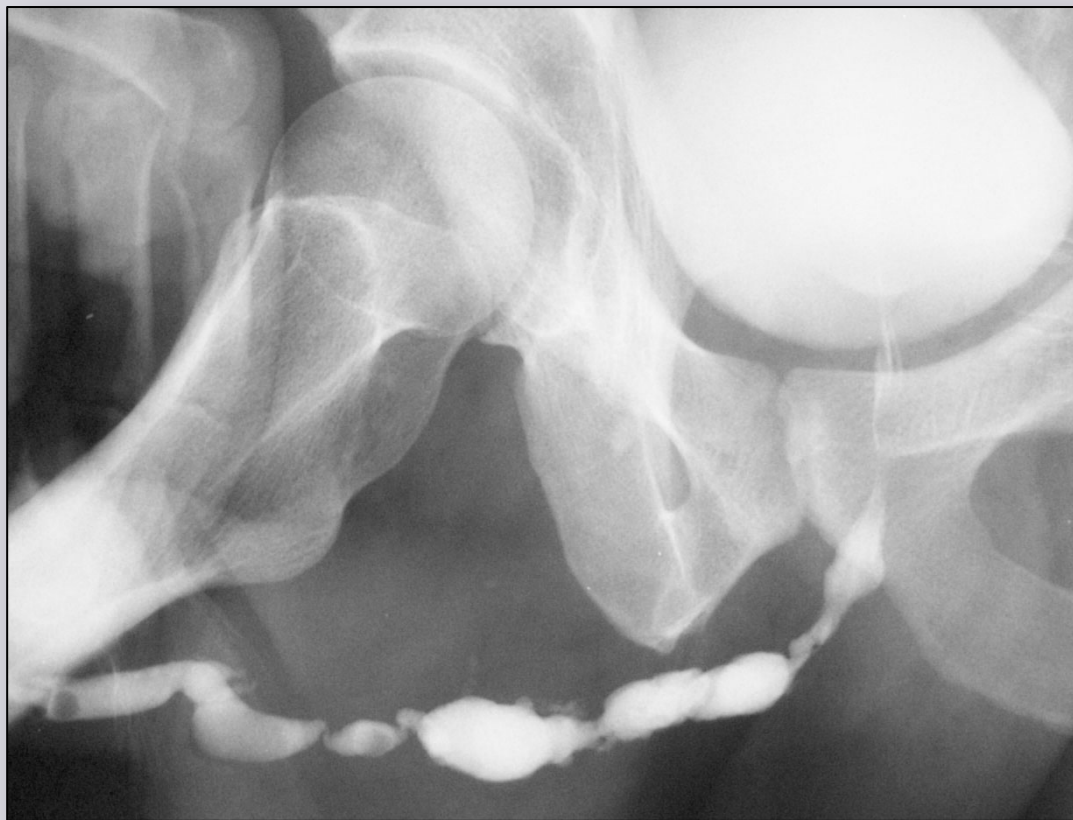
Is tissue engineered oral material fit for use in patient with immunological disorder like Lichen Sclerosus?

Our preliminary experience



Is tissue engineered oral material fit for use in patient requiring two-stage urethral reconstruction?

Our preliminary experience



**Is tissue engineered oral material fit for use in patient requiring
pan-urethral reconstruction?**

e-mail: info@urethralcenter.it

Websites: www.uretra.it
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Our preliminary experience



The use of tissue engineered oral mucosa is not a simple surgical procedure and should be performed only in a Centre of excellence for urethral surgery

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Limitations of this study

- **Short follow-up**

✓ **This material should be used only in Germany**

✓ **The cost is about 4.000,00 to 5.000,00 Euro**

✓ **This material should be used in 48 hours**

Take home message:

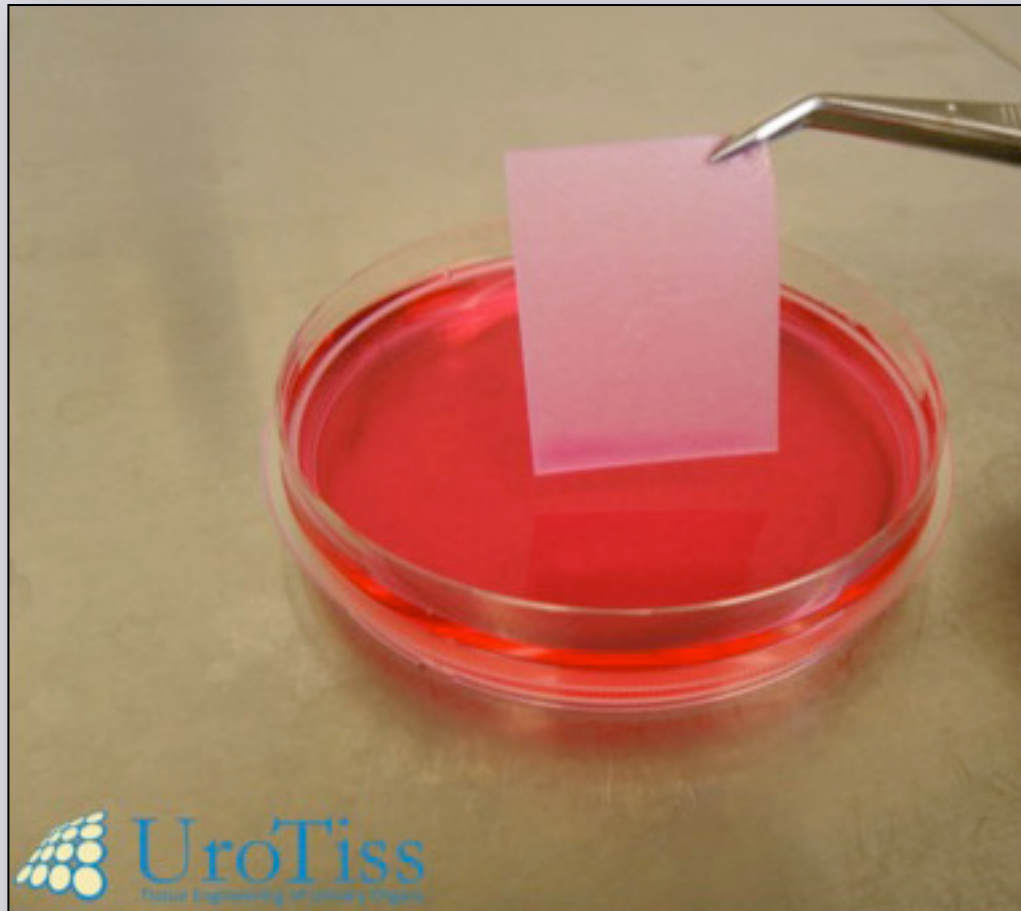
It is not the end line of the long history of urethral reconstruction, but the first step for a new future of urethral surgery.



1987



2014



Next future

2014

For more information about MukoCell manufacturing or if you are interested in developing this technology in your country please contact Dr. Gouya Ram-liebig at:



Email: g.ram-liebig@urotiss.com

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